

# Thermal optimum of photosynthesis is controlled by stomatal conductance and does not acclimate across an urban thermal gradient in six subtropical tree species

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## Abstract

Modelling the response of plants to climate change is limited by our incomplete understanding of the component processes of photosynthesis and their temperature responses within and among species. For  $\geq 20$  individuals, each of six common subtropical tree species occurring across steep urban thermal gradients in Miami, Florida, USA, we determined rates of net photosynthesis ( $A_{\text{net}}$ ), maximum RuBP carboxylation, maximum RuBP regeneration and stomatal conductance, and modelled the optimum temperature ( $T_{\text{opt}}$ ) and process rate of each parameter to address two questions: (1) Do the  $T_{\text{opt}}$  of  $A_{\text{net}}$  ( $T_{\text{optA}}$ ) and the maximum  $A_{\text{net}}$  ( $A_{\text{opt}}$ ) of subtropical trees reflect acclimation to elevated growth temperatures? And (2) What limits  $A_{\text{net}}$  in subtropical trees? Against expectations, we did not find significant acclimation of  $T_{\text{optA}}$ ,  $A_{\text{opt}}$  or the  $T_{\text{opt}}$  of any of the underlying photosynthetic parameters to growth temperature in any of the focal species. Model selection for the single best predictor of  $A_{\text{net}}$  both across leaf temperatures and at  $T_{\text{optA}}$  revealed that the  $A_{\text{net}}$  of most trees was best predicted by stomatal conductance. Our findings are in accord with those of previous studies, especially in the tropics, that have identified stomatal conductance to be the most important factor limiting  $A_{\text{net}}$ , rather than biochemical thermal responses.

## KEYWORDS

acclimation, ecophysiology, gas exchange,  $g_s$ ,  $J_{\text{max}}$ , photosynthetic thermal optimum, thermal gradient,  $V_{\text{cmax}}$

## 1 | INTRODUCTION

Dynamic global vegetation models and earth system models simulate the effects of future climate change on major biogeochemical cycles. These models depend on an accurate understanding of the temperature responses of the component processes of photosynthesis within and among species and across space and time (Kumarathunge et al., 2019; Lin et al., 2012). Currently, models are limited by a poor representation of the potential acclimation of the underlying components of net

photosynthesis ( $A_{\text{net}}$ ) to increased temperature, especially in tropical plants (Lombardozzi et al., 2015; Mercado et al., 2018; Slot et al., 2021; N. G. Smith & Dukes, 2013), due to a sparsity of empirical data characterizing temperature responses (Slot & Winter, 2017a). Earth's atmosphere has already warmed by  $\geq 1^\circ\text{C}$  since 1850–1900 and is predicted to warm by an additional 1.0–5.7°C this century, depending on emissions scenarios (IPCC, 2021). In the face of such rapid climate change, there is an imminent need to understand the global acclimation response of  $A_{\text{net}}$  to rising temperatures.

Photosynthesis has long been recognized as a temperature-sensitive process (Berry & Bjorkman, 1980; Decker, 1959; Hew et al., 1969; Mooney et al., 1978; Pearcy, 1977).  $A_{\text{net}}$  rapidly increases with leaf temperature up to a thermal optimum ( $T_{\text{optA}}$ ) before rapidly decreasing again at higher leaf temperatures (Cunningham & Read, 2003; Säll & Pettersson, 1994). There is large variability in  $T_{\text{optA}}$  and in  $A_{\text{net}}$  at  $T_{\text{optA}}$  ( $A_{\text{opt}}$ ) as a result of both adaptation and individual-level responses to environmental conditions, that is, acclimation (Berry & Bjorkman, 1980; Hikosaka et al., 2005; Lin et al., 2012; Way & Yamori, 2014).

Recent investigations are addressing the challenges surrounding the accurate prediction of both  $T_{\text{optA}}$  and  $A_{\text{opt}}$  acclimation responses to temperature, which stem from an incomplete understanding of the temperature responses—especially in low-latitude tropical species—of the component processes underlying variability in  $T_{\text{optA}}$  and  $A_{\text{opt}}$ , including biochemical, respiratory and stomatal processes (Dusenge & Way, 2017; Hernández et al., 2020; Kattge & Knorr, 2007; Lin et al., 2012; Rogers et al., 2017; Slot & Winter, 2017a).

The Farquhar et al. (1980) model of C3 photosynthesis expresses  $A_{\text{net}}$  in terms of its two major limiting biochemical process rates: the maximum rate of RuBP carboxylation ( $V_{\text{cmax}}$ ) and the maximum rate of RuBP regeneration, or the maximum rate of electron transport ( $J_{\text{max}}$ ). The Farquhar model of photosynthesis is a powerful tool for global change ecology because it mechanistically represents the effects of elevated atmospheric ( $\text{CO}_2$ ) and temperature on the two major biochemical components impacting  $A_{\text{net}}$  (Medlyn et al., 2002). Across increasing leaf temperatures, both  $V_{\text{cmax}}$  and  $J_{\text{max}}$  either increase exponentially or follow a peaked Arrhenius function (Medlyn et al., 2002). Under current ambient ( $\text{CO}_2$ ) and close to a given plant's  $T_{\text{optA}}$ ,  $A_{\text{net}}$  is typically limited by  $V_{\text{cmax}}$  (Sage & Kubien, 2007). However, stomatal conductance ( $g_s$ ), which determines the availability of  $\text{CO}_2$  for photosynthesis and, therefore, exerts control over the achieved carboxylation and electron transport rates in C3 photosynthesis, may be of greater importance than biochemical processes in limiting  $A_{\text{net}}$ , especially in tropical (Doughty & Goulden, 2008; Hernández et al., 2020; Slot & Winter, 2017a, 2017b; Tan et al., 2017) and Mediterranean climates (Sperlich et al., 2019), and in taller, mature trees (Juárez-López et al., 2008). The temperature response of  $g_s$  follows a similar peaked temperature response to  $A_{\text{net}}$  (June et al., 2004), wherein stomata are open within a favourable temperature range, but are closed at colder temperatures and at supra-optimal temperatures associated with increased leaf-to-air vapour pressure deficit (VPD) (Peak & Mott, 2011). However, when moisture is not limiting,  $g_s$  may increase at high temperatures to regulate leaf temperatures (Feller, 2006; Reynolds-Henne et al., 2010), although decreases in  $A_{\text{net}}$  are still observed due to Rubisco inhibition (Crafts-Brandner & Salvucci, 2000; Salvucci & Crafts-Brandner, 2004) and increasing respiration in the light (Silim et al., 2010).

Major patterns of how the temperature responses of these underlying processes vary *interspecifically* across some biomes and plant functional types are relatively well understood (Crous et al., 2022; Kattge & Knorr, 2007; Kumarathunge et al., 2019),

but *intraspecific* responses of  $A_{\text{net}}$  and its mechanistic underpinnings to increased temperature (and increased  $\text{CO}_2$ ), and changes in water availability and humidity), that is, acclimation, remains poorly characterized, especially at low latitudes. This lack of information about intraspecific patterns can hinder the parameterization of global vegetation models (Crous et al., 2022; Kumarathunge et al., 2019) and is especially concerning considering that many tropical species may exhibit weak acclimation of  $A_{\text{net}}$  to warming (Crous et al., 2022; Dusenge, Wittermann, et al., 2021).

Multiple studies of seasonal temperature acclimation in plants have identified a decrease in the ratio of  $J_{\text{max}}:V_{\text{cmax}}$  with increasing ambient temperatures (Atkin et al., 2006; Dusenge et al., 2020; Kumarathunge et al., 2019; Lin et al., 2013; Stefanski et al., 2020). This relationship is associated with either an increase in  $V_{\text{cmax}}$  (Lin et al., 2013) or with a change in the relative amounts of relevant photosynthetic proteins associated with each process rate (Onoda et al., 2005), and may serve to counteract increased photorespiration at high temperatures (Kattge & Knorr, 2007; Kumarathunge et al., 2019; N. G. Smith & Keenan, 2020). A result of decreasing  $J_{\text{max}}:V_{\text{cmax}}$  at high temperatures is that  $A_{\text{net}}$  is less limited by RuBP carboxylation, and  $T_{\text{optA}}$  increases (Crous et al., 2022; Dusenge et al., 2020). Indeed, studies of experimentally warmed plants in temperate and boreal systems have found declines of  $J_{\text{max}}:V_{\text{cmax}}$  with increased growth temperatures of 3–4°C are associated with significant increases in  $T_{\text{optA}}$  (Hara et al., 2021; Kumarathunge et al., 2019; Sendall et al., 2015), an acclimation effect that may sometimes be sufficient to maintain  $A_{\text{net}}$  (Stefanski et al., 2020).

Similarly, experimental warming studies on tropical tree seedlings and saplings have found  $A_{\text{net}}$  to be increasingly limited by  $J_{\text{max}}$  under hotter growth temperatures (Slot & Winter, 2017c; Vårhammar et al., 2015), especially in early successional species (Slot & Winter, 2018). Furthermore, these studies have found partial to full acclimation of  $T_{\text{optA}}$  to increased growth temperatures, that is, increases in growth temperature are equal to increases in  $T_{\text{optA}}$ , but in some cases at a cost to  $A_{\text{opt}}$  (Fauset et al., 2019; Slot et al., 2021; Slot & Winter, 2017c, 2018). However,  $g_s$  may be a greater limit to  $A_{\text{net}}$  as growth temperature increases, especially when water is limiting and leaf-to-air VPD is high, as often occurs in the upper canopies of mature trees (Fauset et al., 2019). Indeed, at constant absolute humidity, VPD increases exponentially with temperature, so rising temperatures are typically accompanied by rising VPD. Finally, respiration, which typically increases exponentially with leaf temperature, has been shown to acclimate and thus may also play an important role in the overall observed acclimation of  $A_{\text{net}}$  and  $T_{\text{optA}}$  (Atkin & Tjoelker, 2003; Crous et al., 2022). More studies, especially with species from the subtropics and tropics, are needed to better characterize the long-term acclimation of photosynthetic parameters to increased growth temperatures (Dusenge & Way, 2017; Slot et al., 2021), which will improve the parameterization of global dynamic vegetation models.

Testing for the acclimation of mature trees across their development and lifetimes is logistically challenging and time consuming. 'Accidental experiments' (HilleRisLambers et al., 2013),

or pseudo-experimental setups created inadvertently by humans, can provide opportunities to address questions regarding global change while avoiding some of the challenges of true experiments. For example, urban areas are often significantly warmer than surrounding rural areas, primarily due to the prevalence of man-made surfaces that absorb solar radiation and release heat. This urban heat island effect creates thermal gradients that are exceptionally strong and steep, even over short distances within urban areas (e.g., 5°C difference over 10–100 s of metres vs. natural gradients of ~5°C per 1000 m elevation or ~5°C per 1 000 000 m latitude), making them useful settings for studying the acclimation of trees to different temperatures (Carreiro & Tripler, 2005; Esperon-Rodriguez et al., 2020; Hara et al., 2021; Kullberg & Feeley, 2022; Meineke et al., 2016; Youngsteadt et al., 2015). Other environmental factors such as CO<sub>2</sub>, ozone and deposited nitrogen may covary with temperature on urban-to-rural gradients, but working along intra-urban temperature gradients while excluding rural areas may help to avoid significant impacts of potential confounding variables.

In this study, we compared the acclimation of photosynthetic parameters to different growth temperatures for six common tree species planted across thermal gradients in Miami, Florida, USA, urban heat island. We determined the rates of net photosynthesis ( $A_{\text{net}}$ ), maximum RuBP carboxylation ( $V_{\text{cmax}}$ ), maximum RuBP regeneration ( $J_{\text{max}}$ ) and stomatal conductance ( $g_s$ ) at a range of leaf temperatures, and modelled the thermal optimum ( $T_{\text{opt}}$ ) and optimum process rate ( $k_{\text{opt}}$ ) of each parameter to address two research questions: (1) Do these subtropical tree species acclimate their  $T_{\text{optA}}$  and  $A_{\text{opt}}$  to elevated growth temperatures? And (2) what limits  $A_{\text{net}}$  in these focal species, and does the limiting process vary within species based on growth temperature? We predicted that there would be consistent increases in  $T_{\text{optA}}$  and changes in  $A_{\text{opt}}$  for trees growing under hotter temperatures, but that acclimation of  $T_{\text{optA}}$  would be partial, that is, the increase in  $T_{\text{optA}}$  would be smaller than the increase in growth temperature. We also predicted species-specific changes in  $A_{\text{opt}}$  with changes in growth temperature or  $T_{\text{optA}}$ , but the directionality of this relationship should vary and be dependent on the acclimation of the various processes underlying  $A_{\text{net}}$  (Way & Yamori, 2014). Finally, we predicted that  $g_s$  would be a greater limit to the temperature response of  $A_{\text{net}}$  than the biochemical processes, and that  $g_s$  limitation would increase for trees growing in hotter areas.

## 2 | MATERIALS AND METHODS

This study was conducted in the urban and suburban areas of subtropical Miami-Dade County, southeast Florida, USA, taking advantage of the marked spatial variation in ambient temperatures (see below) created through the urban heat island effect. We measured gas exchange rates and estimated associated parameters on the leaves of 121 planted (i.e., not naturally recruited) trees representing six focal species ( $\geq 20$  trees per species), all of which are common and widespread throughout the study area: *Bursera simaruba* (L.) Sarg., *Coccoloba uvifera* (L.) L., *Quercus virginiana* Mill., *Heptapleurum actinophyllum* (Endl.) Lowry

& G.M. Plunkett, *Swietenia mahagoni* (L.) Jacq and *Terminalia catappa* L. The species *B. simaruba*, *C. uvifera*, *Q. virginiana* and *S. mahagoni* are all native to the study area, and the species *H. actinophyllum* and *T. catappa* are exotic (for more information about the study system and species, see Kullberg & Feeley, 2022).

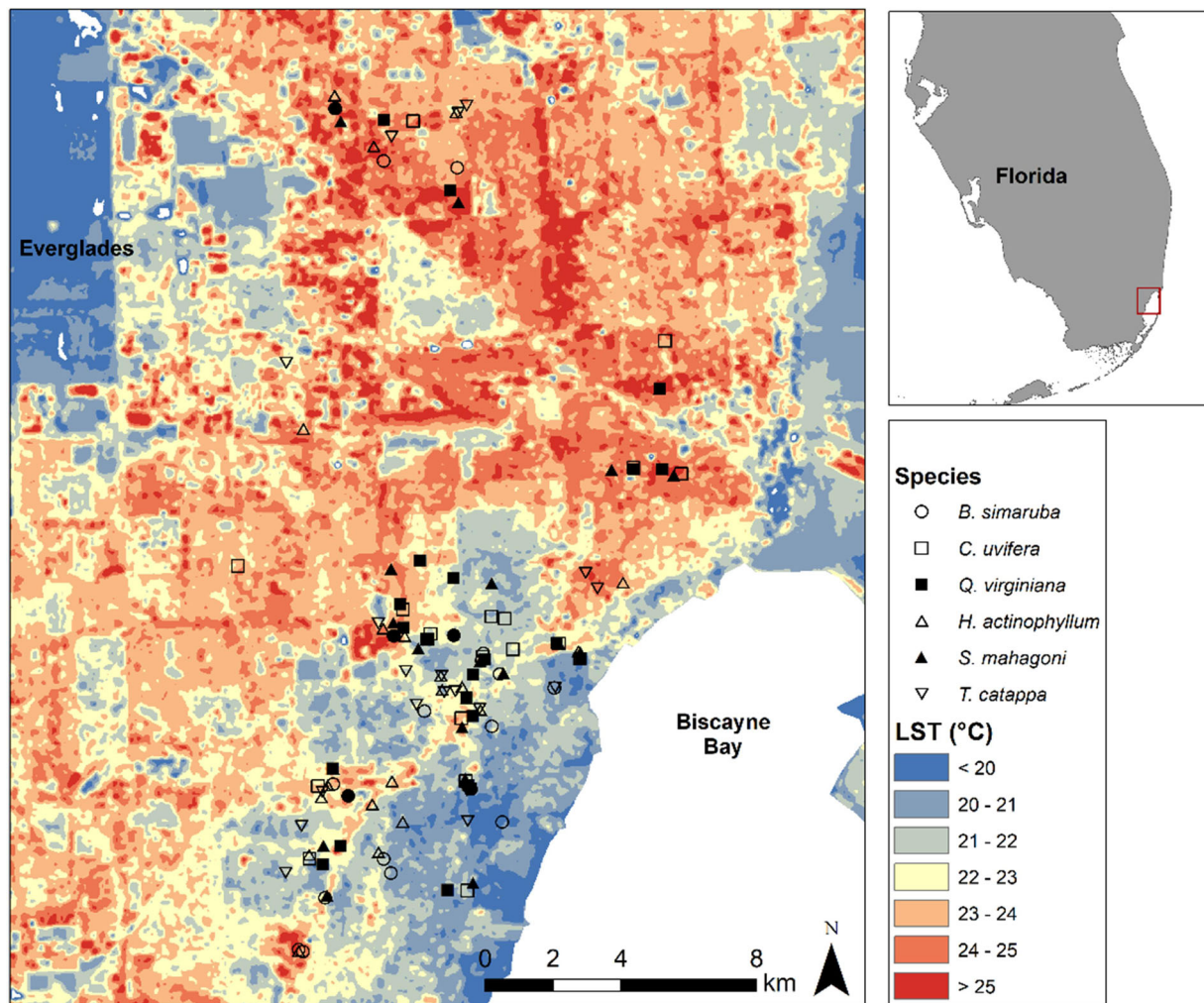
### 2.1 | Sample collection

We selected focal trees to maximize the range of ambient temperatures to which they are exposed, and to capture as much of the gradient along that range as possible, rather than maximizing replication at a few select temperatures. The maximum straight-line distance between conspecific pairs was 25.2 km (Figure 1). Canopies of focal trees had near-total to total sun exposure, and no samples were collected close to tall buildings or structures that could shade the trees. Between June and October 2021, we collected a roughly 1 m long ( $\pm 0.3$  m) sun-exposed branch from the outer canopy of each tree between 08:00 and 09:30 local time (for *H. actinophyllum* we collected individual leaves due to them having thick branches and sufficient petiole length to maintain water transport). We immediately placed the cut branch ends into water and covered the branches and leaves with opaque plastic bags to maintain a dark, humid environment and to prevent wilting. We recorded the percent of ground cover that was impervious to water (i.e., concrete or pavement) within a 5 m radius of each tree and collected two soil cores (up to 15 cm deep when bedrock was deep enough) within 3 m of each tree; soil cores were subsequently mixed to create one aggregate soil sample per tree. Soil samples were air-dried in the laboratory for at least 1 week before analyses.

Upon returning to the laboratory (within 1 h of sample collection), branches were re-cut under water to promote water transport, re-covered with plastic to prevent wilting, and allowed to acclimate to the laboratory conditions for >1 h before connecting to the gas analyser. Connected branches were left to acclimate to ambient air/light and pre-set leaf chamber conditions for at least 30 min before data collection. While it is generally preferable to measure gas exchange on leaves of attached branches, this is not always possible for sun-exposed canopy branches of adult trees. Some previous studies have shown no significant effects of branch excision on photosynthetic parameters (Verryckt et al., 2020), while other studies (Davidson et al., 2022) have shown that afternoon stomatal conductance measurements may be reduced for excised branches in some species—in particular, in latex-producing species the maximum rates of  $A_{\text{net}}$ ,  $V_{\text{cmax}}$  and  $J_{\text{max}}$  may be significantly reduced in excised branches (Santiago & Mulkey, 2003). Since we did not use any latex-produce species in our study and since we are primarily interested in within-species relationships, we expect that our results were not significantly affected by branch excision.

### 2.2 | Gas exchange measurements

To estimate the thermal optima ( $T_{\text{opt}}$ ) and the temperature responses of the underlying biochemical mechanisms limiting net photosynthesis



**FIGURE 1** A map of the study area in the Miami urban heat island, showing land surface temperature (LST) and the location of each focal tree

( $A_{\text{net}}$ ) of individual trees, we measured net  $\text{CO}_2$  assimilation across a range of leaf intercellular  $\text{CO}_2$  concentrations ( $A_C$  curves) nested within a range of leaf temperatures. Specifically, we measured gas exchange rates at 12 different ambient  $\text{CO}_2$  concentrations ranging from 50 to 2000 ppm nested within 10 different leaf temperatures (24°C, 26°C, 28°C, 30°C, 32°C, 34°C, 36°C, 38°C, 40°C and 42°C) using a LiCor LI-6800 portable gas exchange system (LI-COR Inc.). All gas exchange measurements were conducted in a laboratory setting, and all environmental conditions were manipulated only inside the leaf chamber using an LI-6800. The gas exchange rates were measured on one healthy, mature leaf per focal individual (resulting in 120 measurements per tree), which were clamped into the 6 cm<sup>2</sup> aperture leaf chamber if leaf lamina were large enough and otherwise were clamped into the 2 cm<sup>2</sup> aperture leaf chamber. Reference  $\text{CO}_2$  concentrations and leaf temperatures were controlled by the LI-6800 instrument for all measurements, leaf surface irradiance was maintained at 1000  $\mu\text{mol quanta m}^{-2} \text{s}^{-1}$ , and the sample VPD was set to 1.5 kPa (to prevent condensation within the LI-6800 at lower temperatures). Keeping a constant VPD avoids confounding changes in temperature with changes in VPD on stomatal conductance ( $g_s$ ), especially at high

temperatures where stomatal aperture may be independent of  $A_{\text{net}}$  (Urban et al., 2017). To avoid sensor drift, the LI-6800's reference and sample chamber infra-red gas analysers were matched before each measurement. We programmed stability criteria for the instrument to determine when it was appropriate to log a measurement, including the slope of  $A_{\text{net}}$  (slope limit of 0.8 and SD of 1 over 15 s) and  $g_s$  (slope limit of 0.2 and SD of 1 over 15 s). We had the instrument wait at least 2 min between measurements to reach stability (and an additional 4 min when changing leaf temperature) and allowed up to 3 min before logging measurements. In total, it took approximately 4.5 h per leaf to conduct the gas exchange measurements at all 12 reference  $\text{CO}_2$  concentrations at all 10 leaf temperatures, and the same leaf remained within the leaf chamber during the entire protocol.

### 2.3 | Parameter estimation and temperature responses

We estimated the maximum rate of Rubisco carboxylation ( $V_{\text{cmax}}$ ) and the maximum rate of electron transport for RuBP regeneration ( $J_{\text{max}}$ )



with the 'fitaci' function in the 'plantecophys' package (Duursma, 2015) in R (v. 4.1.2) after visually inspecting the data for each  $AC_i$  curve and omitting any measurements that fell outside a reasonable range of values ( $0 \text{ ppm} < C_i < 2500 \text{ ppm}$  and  $-1 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1} < A_{\text{net}} < 50 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ ) or varied greatly from an otherwise visually discernible curve, a sign of the LI-6800 not reaching stability before logging a measurement. Unfortunately, the first point taken for the  $AC_i$  curves (at ambient  $[\text{CO}_2]$  ( $C_a$ ) = 400 ppm) were often discarded since extra time was generally required to reach stability after a change in leaf temperature. We estimated  $V_{\text{cmax}}$  and  $J_{\text{max}}$  for each measured leaf temperature instead of using the default correction to 25°C. We inspected every modelled  $AC_i$  curve visually and omitted curves with less than 5 points.

To estimate the  $T_{\text{opt}}$  of  $V_{\text{cmax}}$  ( $T_{\text{opt}V}$ ) and  $J_{\text{max}}$  ( $T_{\text{opt}J}$ ) for each individual, we fit the following peaked Arrhenius equation with the 'nls' function in R according to Medlyn et al. (2002):

$$f(T_{\text{leaf}}) = k_{\text{opt}} \frac{H_d \times e^{\left(\frac{H_a(T_{\text{leaf}} - T_{\text{opt}})}{T_{\text{leaf}} \times R \times T_{\text{opt}}}\right)}}{H_d - H_a \left(1 - e^{\left(\frac{H_d(T_{\text{leaf}} - T_{\text{opt}})}{T_{\text{leaf}} \times R \times T_{\text{opt}}}\right)}\right)}, \quad (1)$$

where  $T_{\text{leaf}}$  is leaf temperature (in K),  $k_{\text{opt}}$  is the process rate of either  $V_{\text{cmax}}$  ( $V_{\text{opt}}$ ) or  $J_{\text{max}}$  ( $J_{\text{opt}}$ ) at the respective  $T_{\text{opt}}$  (in  $\mu\text{mol m}^{-2} \text{ s}^{-1}$ ) of each;  $H_a$  is the activation energy of the Arrhenius function or the exponential rise before  $T_{\text{opt}}$ ;  $R$  is the universal gas constant ( $0.008314 \text{ kJ K}^{-1} \text{ mol}^{-1}$ ); and  $H_d$  is the deactivation energy, or the decrease after  $T_{\text{opt}}$ , fixed at  $400 \text{ kJ mol}^{-1}$  to avoid overparameterization of the models. We set  $H_d$  to  $400 \text{ kJ mol}^{-1}$  after determining that it explained on average, a greater proportion of variance in the temperature response of both  $V_{\text{cmax}}$  and  $J_{\text{max}}$  than the more standard  $200 \text{ kJ mol}^{-1}$  (Medlyn et al., 2002) in most of our study species (Supporting Information: Figure S1; setting  $H_d$  to  $400 \text{ kJ mol}^{-1}$  tended to allow the model to better approximate the high observed maximum process rates of  $V_{\text{cmax}}$  and  $J_{\text{max}}$ ).

Starting values for estimating parameters were set as  $H_a = 100 \text{ kJ mol}^{-1}$ ,  $k_{\text{opt}}$  = the maximum value of either  $V_{\text{cmax}}$  or  $J_{\text{max}}$  and  $T_{\text{opt}}$  = the leaf temperature corresponding to the maximum value of either  $V_{\text{cmax}}$  or  $J_{\text{max}}$ . We estimated  $V_{\text{cmax}25}$  and  $J_{\text{max}25}$  as the fitted peaked Arrhenius model predictions of each process rate at 25°C to evaluate if  $J_{\text{max}25} : V_{\text{cmax}25}$  (i.e., the ratio between these two processes) decreased with increasing growth temperature as observed in previous studies (Kattge & Knorr, 2007; Kumarathunge et al., 2019; N. G. Smith & Dukes, 2013). We also estimated the ratio of  $J_{\text{max}}$  and  $V_{\text{cmax}}$  at  $T_{\text{opt}A}$  ( $J_{\text{max}T_{\text{opt}}} : V_{\text{cmax}T_{\text{opt}}}$ ). Additionally, we calculated the entropy term ( $\Delta S$ ) of  $V_{\text{cmax}}$  and  $J_{\text{max}}$  according to Medlyn et al. (2002) and Slot et al. (2021) as:

$$\Delta S = \frac{H_d}{T_{\text{opt}}} + R \ln\left(\frac{H_a}{H_d - H_a}\right). \quad (2)$$

We examined the temperature response of the activation state of Rubisco following the methods of Sage et al. (2008), wherein the slope of the linear regression of observed  $A_{\text{net}}$  values at low  $C_i$  ( $\leq 200 \text{ ppm}$ , where Rubisco activase should not be limited

by ATP supply) may be compared to the initial slope of the modelled  $AC_i$  curve, which follows the assumption that Rubisco is fully activated:

$$\text{Initial slope} = \frac{V_{\text{cmax}}}{\left(\Gamma^* + K_C \left(1 + \frac{O}{K_O}\right)\right)}, \quad (3)$$

where  $\Gamma^*$  is the  $\text{CO}_2$  compensation point in the absence of dark respiration and  $K_C$  and  $K_O$  are the Michaelis–Menten constants for  $\text{CO}_2$  and  $\text{O}_2$ , respectively, which are both temperature-dependent parameters calculated per treatment temperature according to Bernacchi et al. (2001).  $O$  is the concentration of  $\text{O}_2$  in the chloroplast stroma, assumed to be 210 mbar. By comparing the observed initial slope against the modelled slope at low  $C_i$  where electron transport is not limiting, we tested the model's assumption that Rubisco is fully activated at a given leaf treatment temperature.

The temperature response of observed versus modelled Rubisco activation was then modelled by fitting generalized additive models (GAMs) with the 'mgcv' package (Wood, 2017) in R, using cubic regression splines fit with restricted maximum likelihood, as in Slot et al. (2021). Rubisco activation was determined to be significantly reduced where 1.96 times the standard error of the observed and modelled GAMs did not overlap.

$A_{\text{net}}$  and  $g_s$  were modelled as functions of temperature following the model presented in June et al. (2004):

$$k(T_{\text{leaf}}) = k_{\text{opt}} \times e^{-\left(\frac{T_{\text{leaf}} - T_{\text{opt}}}{\Omega}\right)^2}, \quad (4)$$

where  $T_{\text{leaf}}$  is leaf temperature,  $k_{\text{opt}}$  is the process rate of either assimilation ( $A_{\text{opt}}$ ) or  $g_s$  ( $g_{\text{opt}}$ ), and  $\Omega$  is defined as the difference between the temperatures above and below  $T_{\text{opt}}$  at which the process rate is reduced by  $\sim 37\%$  from  $k_{\text{opt}}$ . Since many measurements of  $A_{\text{net}}$  had to be discarded when  $C_a$  was set to 400 ppm due to insufficient acclimation time to new  $T_{\text{leaf}}$ ,  $A_{\text{net}}$  was extracted from each  $AC_i$  curve as the net C assimilation rate modelled at the respective intercellular  $\text{CO}_2$  concentration ( $C_i$ , in ppm) measured when  $C_a$  was set to 400 ppm. While our reported  $A_{\text{net}}$  values are, therefore, estimations based on modelled  $AC_i$  curves, this method allowed us to integrate all the collected photosynthesis data into the estimates of  $A_{\text{net}}$ .  $g_s$  was calculated as the average measured  $g_s$  when  $C_i$  was  $\pm 150 \text{ ppm}$  of the  $C_i$  of extracted  $A_{\text{net}}$ .

Stomatal limitation ( $l$ ) was determined according to (Farquhar & Sharkey, 1982), Slot and Winter (2017b), and Hernández et al. (2020) as:

$$l = 1 - \frac{A_{\text{obs}}}{A_{\text{inf}g_s}}, \quad (5)$$

where  $A_{\text{obs}}$  is the observed rate of  $A_{\text{net}}$  and  $A_{\text{inf}g_s}$  is the rate of  $A_{\text{net}}$  if there were no stomatal limitation, determined according to the Farquhar et al. (1980) model of C3 photosynthesis as:

$$V_{\text{cmax}} = \frac{A_g(C_i + K_C)}{C_i - \Gamma^*}, \quad (6)$$

where  $C_i$  is set to  $C_a$  (around 400 ppm) and  $A_g$  is the rate of light-saturated photosynthesis plus respiration in the light (estimated as  $0.015 \times V_{cmax}$ ).  $A_{inf\ g_s}$  was then determined as  $A_g$  minus respiration in the light.

## 2.4 | Growth temperatures

We estimated the relative ambient temperature at the location of each focal tree based on the land surface temperature (LST) measured with remotely sensed imagery. To create a map of LST, we used a Landsat 8 (Level-1, Collection-1) image taken on 5 January 2021, at 15:50, downloaded from the United States Geological Survey (USGS) EarthExplorer (Landsat Missions). This was the clearest image over the study area for the year before sample collection based on a filter to only include images in the search with <40% cloud cover and visual inspection of images. The Landsat 8 thermal band has 100-m resolution, but LST is calculated at 30-m resolution due to a correction for emissivity based 30-m resolution data on normalized difference vegetation index (NDVI). We calculated LST following USGS protocol (Jeevalakshmi et al., 2017; Kullberg & Feeley, 2022). We then determined the LST at each tree location using the 'extract' function of the 'raster' package in R. In a previous publication, we showed that Landsat 8-derived LST in Miami are highly correlated ( $r \geq 0.8$ ) across months (October to May) and years (2016–2021), indicating that the relative temperature patterns in Miami are consistent across time and seasons, although the absolute temperatures may vary (Kullberg & Feeley, 2022).

## 2.5 | Soil parameters

We measured the pH of each soil sample (one aggregate sample per tree location) using an Extech ExStik II pH/conductivity probe. Samples were prepared by mixing 10 g of dried, crushed soil with 10 ml deionized water, which was then allowed to rest for 30 min before inserting the pH probe into the sample and waiting for the reading to stabilize. The pH probe was calibrated before each use with buffer solutions of pH 4, 7 and 10. For soil nitrogen (N), phosphorus (P) and potassium (K), we used the LaMotte NPK Soil Kit (3-5880) and followed standardized protocols to categorize nutrient concentrations into six categories: trace, trace/low, low, low/medium, medium/high, or high.

## 2.6 | Hypothesis testing

To test our first hypothesis that subtropical trees have a limited ability to acclimate their  $T_{optA}$  and  $A_{opt}$  to elevated temperatures across thermal gradients, we constructed linear least squares regressions comparing the  $T_{optA}$  and the  $A_{opt}$  of each focal individual to LST or % impervious cover. We further explored potential relationships between underlying biochemical and physiological

processes to environmental factors and photosynthetic optima. We tested for relationships between the  $T_{opt}$ s and  $k_{opt}$ s of  $V_{cmax}$ ,  $J_{max}$  or  $g_s$  and LST, % impervious cover,  $T_{optA}$  or  $A_{opt}$ . We also examined relationships for  $J_{maxTopT}:V_{cmaxTopT}$ ,  $J_{max25}:V_{cmax25}$ , and the slopes of  $J_{max}:V_{cmax}$  or  $I$  over leaf temperature, compared to LST, % impervious cover,  $T_{optA}$  or  $A_{opt}$ .

To address the possibility that the acclimation responses of  $T_{optA}$  and  $A_{opt}$  covary with environmental factors other than temperature, we constructed linear mixed models (LMMs) using the 'lmerTest' package in R (Kuznetsova et al., 2017). We set the full models with the response variable as either  $T_{optA}$  or  $A_{opt}$ , species as a random intercept and the environmental factors of LST, % impervious cover (as a rough proxy for water availability), soil soluble N, P and K, and soil pH as fixed effects. Next, we used the 'step' function to perform backward stepwise model selection given the full model and without reducing random effects (i.e., species effects).

To further explore which parameter ( $V_{cmax}$ ,  $J_{max}$  or  $g_s$ ) is most limiting to  $A_{net}$ , we used the 'leaps' package in R (Lumley, 2004) to search exhaustively for the best single-predictor model of  $A_{net}$  for each tree using a branch-and-bound algorithm and using  $R^2$ , adjusted  $R^2$  and Mallows'  $C_p$  as selection criteria (Slot & Winter, 2017a). We repeated this algorithm twice per individual, once using the assimilation at all leaf temperatures tested, and again using just the leaf temperatures that are greater than  $T_{optA}$  of that individual. While this model selection method does not directly account for mechanistic dynamics between  $g_s$ ,  $V_{cmax}$  and  $J_{max}$  across temperature, it allowed us to compare the overall limitations to photosynthesis across leaf temperatures with the high-temperature limitations to photosynthesis.

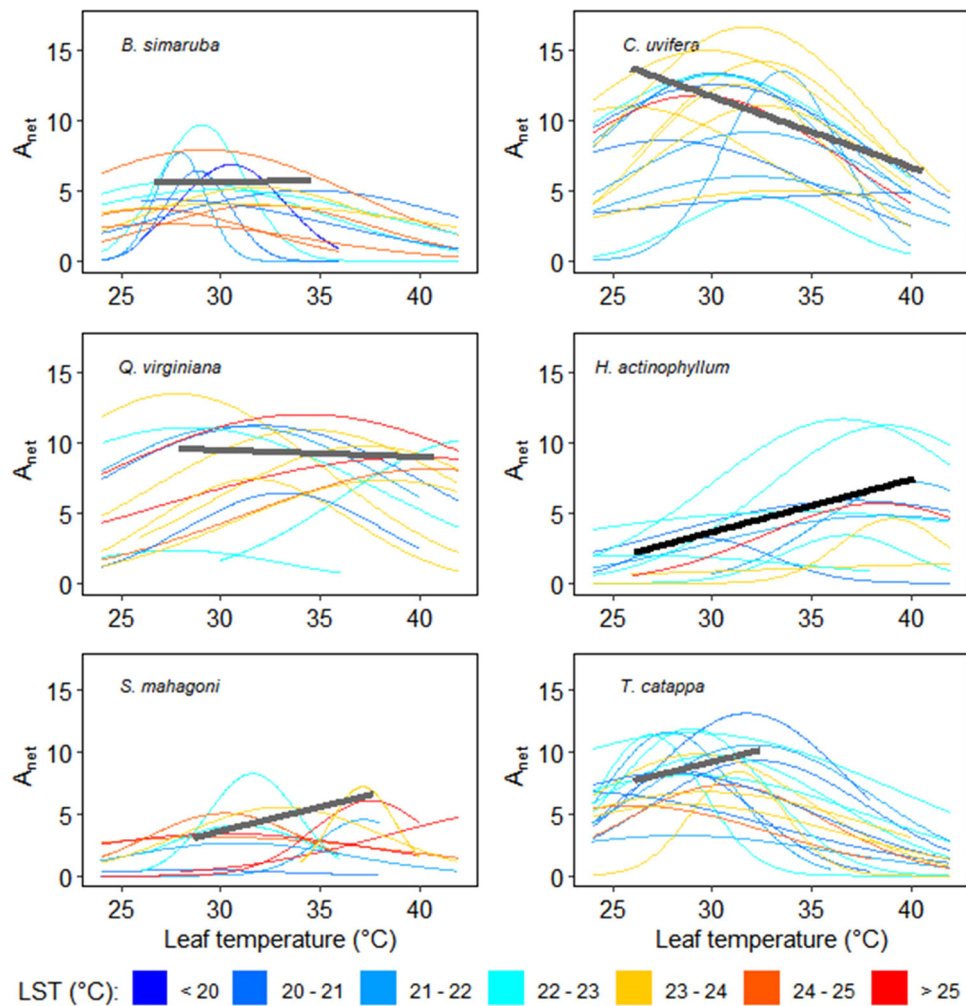
Once we identified the single best predictor of  $A_{net}$  for each tree, we tested whether the difference in the best predictor between the individuals of a given species was associated with differences in either  $T_{optA}$  or the  $R^2$  of the predictive model using analysis of variance (ANOVA). Finally, we compared the proportion of individuals per species that were best predicted by each variable using ANOVA, to see if there was a single variable that consistently predicted  $A_{net}$  across species.

## 3 | RESULTS

Gas exchange measurements on the 121 individuals across 10 leaf measurement temperatures per tree resulted in 1152 successfully modelled  $AC_i$  curves. A summary of the linear relationships between the temperature responses of different photosynthesis parameters to growth temperature and % impervious cover and to each other can be found in Supporting Information: Tables S1 and S2, respectively.

### 3.1 | $k_{opt}-T_{opt}$ relationships by species

$A_{opt}$  was significantly positively correlated with  $T_{optA}$  in one of the study species, *H. actinophyllum* (Figure 2), although this species had



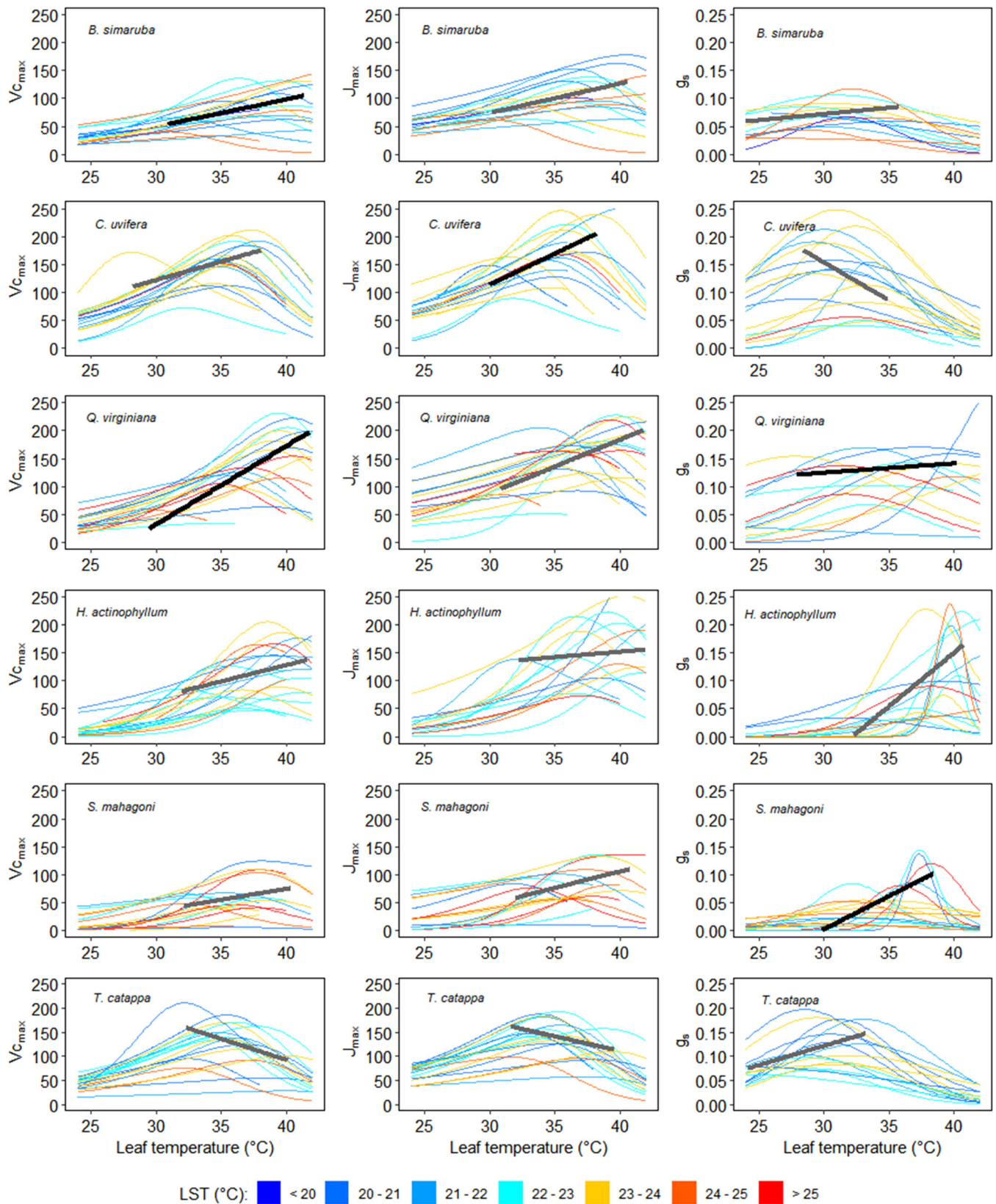
**FIGURE 2** Modelled temperature response curves of  $A_{\text{net}}$  ( $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ ), where each panel is a separate study species, and each curve is one tree. Curves were modelled with the 'nls' package in R using Equation 4. The colour of each curve indicates the relative growth temperature of the tree based on land surface temperature (LST). Black and grey straight lines represent the significant and nonsignificant, respectively, linear regressions of the optimum rate of photosynthesis ( $A_{\text{opt}}$ ) over the thermal optimum of photosynthesis ( $T_{\text{optA}}$ ). The distribution of pseudo- $R^2$  values for the 'nls' models is reported by species in Supporting Information: Figure S3.

the greatest variability in the goodness of fit of its modelled  $A_{\text{net}}$  temperature response curves (Supporting Information: Figure S3). Indeed, the goodness-of-fit of the temperature response curves of both  $A_{\text{net}}$  and  $g_s$  were highly variable across individuals of most species (Supporting Information: Figures S3 and S6).  $V_{\text{opt}}$  was significantly positively correlated with  $T_{\text{optV}}$  in *B. simaruba* ( $p = 0.03$ , adj.  $R^2 = 0.30$ ) and *Q. virginiana* ( $p = 0.002$ , adj.  $R^2 = 0.44$ ; Figure 3), the two study species with the greatest goodness of fit of their  $V_{\text{cmax}}$  temperature response curves (Supporting Information: Figure S4). Also, the activation energy of  $V_{\text{cmax}}$  ( $H_{\text{AV}}$ ) was negatively related to  $T_{\text{optV}}$  in four species, including *B. simaruba* ( $p = 0.001$ , adj.  $R^2 = 0.48$ ), *C. uvifera* ( $p < 0.001$ , adj.  $R^2 = 0.51$ ), *H. actinophyllum* ( $p = 0.03$ , adj.  $R^2 = 0.17$ ) and *T. catappa* ( $p = 0.002$ , adj.  $R^2 = 0.41$ ; Supporting Information: Figure S7A); and  $H_{\text{AV}}$  was positively related to  $V_{\text{opt}}$  in *T. catappa* ( $p = 0.001$ , adj.  $R^2 = 0.44$ ; Supporting Information: Figure S7B).  $J_{\text{opt}}$  was significantly positively related to  $T_{\text{optJ}}$  in

*C. uvifera* ( $p = 0.03$ , adj.  $R^2 = 0.23$ ; Figure 3). The activation energy of  $J_{\text{max}}$  ( $H_{\text{AJ}}$ ) was negatively related to  $T_{\text{optJ}}$  in three species: *B. simaruba* ( $p = 0.045$ , adj.  $R^2 = 0.18$ ), *C. uvifera* ( $p = 0.007$ , adj.  $R^2 = 0.34$ ) and *T. catappa* ( $p = 0.04$ , adj.  $R^2 = 0.19$ ; Supporting Information: Figure S7C), and  $H_{\text{AJ}}$  was positively related to  $T_{\text{optJ}}$  in *S. mahagoni* ( $p = 0.03$ , adj.  $R^2 = 0.22$ ; Supporting Information: Figure S7C).  $H_{\text{AJ}}$  was positively related to  $J_{\text{opt}}$  for one species, *T. catappa* ( $p = 0.002$ , adj.  $R^2 = 0.42$ ; Supporting Information: Figure S7D).  $g_{\text{opt}}$  was significantly positively related to  $T_{\text{optg}}$  in *Q. virginiana* ( $p = 0.01$ , adj.  $R^2 = 0.37$ ) and *S. mahagoni* ( $p = 0.003$ , adj.  $R^2 = 0.41$ ; Figure 3).

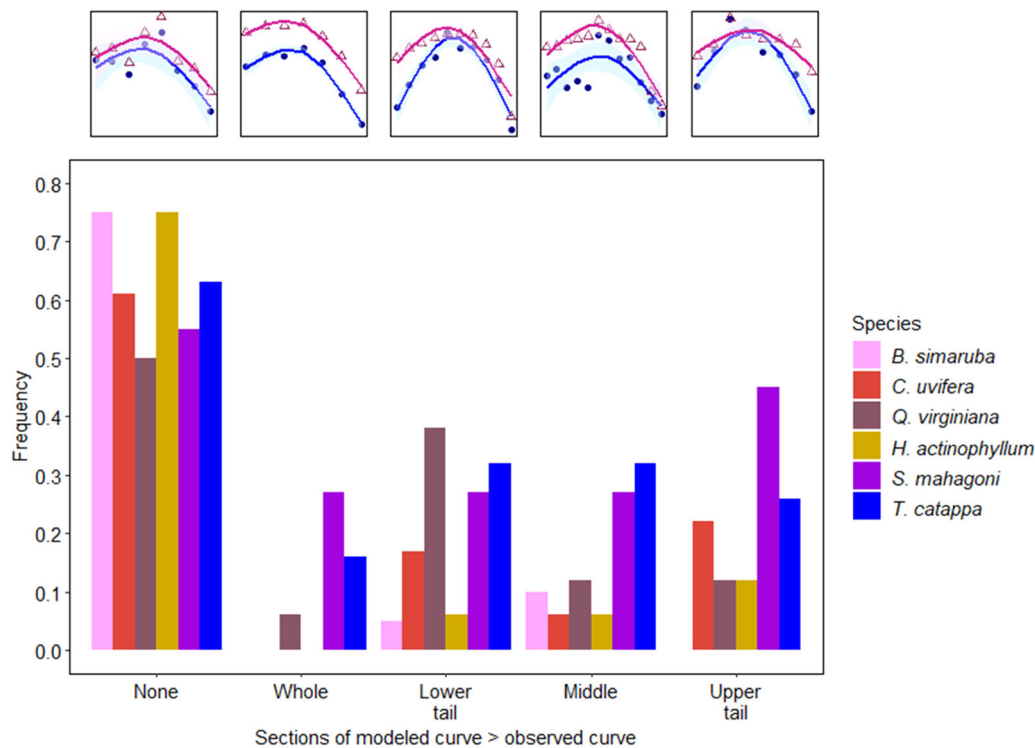
In most trees, the activation state of Rubisco did not limit  $A_{\text{net}}$  at any temperature—observed slopes were not significantly reduced relative to modelled slopes, and the fitted curves overlapped (ANOVA and Tukey's HSD,  $p < 0.001$  between frequency of individuals with complete overlap and frequency of all other categories of curve noncoincidence; Figure 4). However, 10%–25%





**FIGURE 3** Modelled temperature response curves of  $V_{cmax}$  and  $J_{max}$  ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) (Equation 1) and of  $g_s$  ( $\text{mol m}^{-2} \text{s}^{-1}$ ) (Equation 2). Each row of plots is a single study species, and within each panel, each curve is one tree. The colour of each curve indicates the relative growth temperature based on land surface temperature (LST). Black and grey straight lines represent the significant and nonsignificant, respectively, linear regressions of the optimum process rate ( $k_{opt}$ ) over the thermal optimum of the given process ( $T_{optk}$ ). The distribution of pseudo- $R^2$  values for the 'nls' models is reported by parameter and species in Supporting Information: Figures S4–S6.





**FIGURE 4** The sections of the temperature response curve of Rubisco activation that are significantly greater when calculated based on the modelled initial slope of the  $AC_i$  curve (i.e., using  $V_{cmax}$ ) than when calculated based on the observed initial slope of the  $AC_i$  curve (i.e., using raw data points at  $C_i < 200$  ppm). The top panels show exemplar figures of each category of curve noncoincidence; the modelled curve of each individual is red, the observed curve is blue, and the shaded regions are the standard error of the generalized additive model. In the main bottom panel, curve noncoincidence is summarized by individual and grouped by species.

of individuals in all species except *B. simaruba* did have reduced Rubisco activation at high treatment temperatures. *Q. virginiana* and *T. catappa* also had reduced Rubisco activation at relatively low temperatures in 30%–40% of individuals.

### 3.2 | $T_{opt}$ and $k_{opt}$ relationships to growth temperature (LST) and other environmental factors

Against expectations,  $T_{optA}$  did not have a significant positive relationship to LST in any of the six study species (Figure 5a). Similarly, no species showed a significant relationship between  $A_{opt}$  and LST (Figure 5b). There were also no significant single-species relationships between either  $T_{optA}$  or  $A_{opt}$  and % impervious cover (Figure 5c,d).

Similarly, there were also almost no significant single-species relationships between LST and any of the other photosynthesis variables of interest ( $T_{optV}$ ,  $V_{opt}$ ,  $T_{optJ}$ ,  $J_{opt}$ ,  $T_{optg}$ ,  $J_{max}$ : $V_{cmax}$ ,  $J_{max25}$ : $V_{cmax25}$ ,  $J_{maxTopt}$ : $V_{cmaxTopt}$ ,  $g_{opt}$ ), the one exception being a significant negative relationship between  $T_{optV}$  and LST and between  $T_{optJ}$  and LST for *Q. virginiana* (Supporting Information: Table S5). Additionally, none of the species showed notable relationships between % impervious cover and the photosynthesis variables (Supporting Information: Table S5).

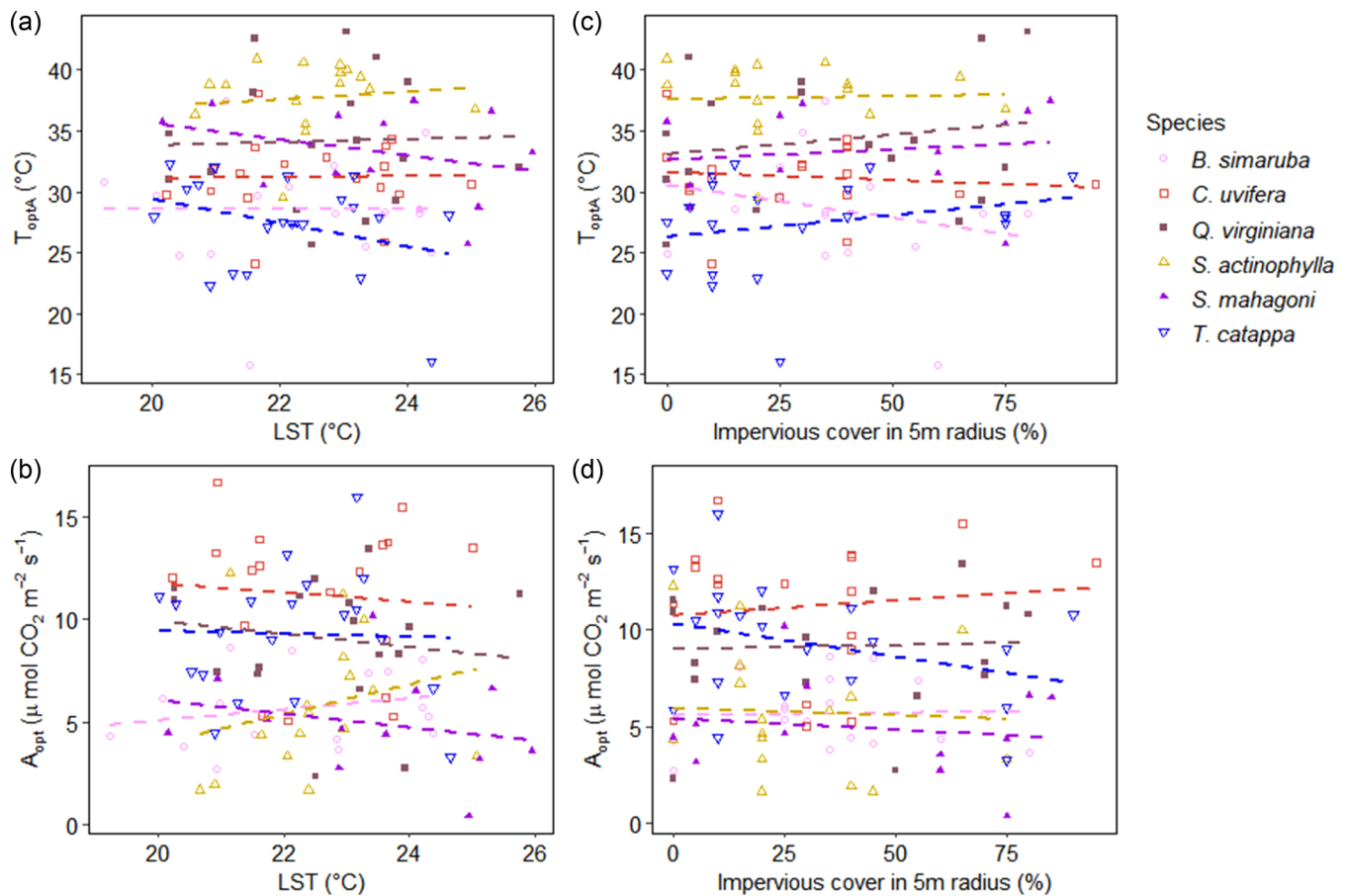
When all environmental variables were used to construct a stepwise LMM predicting  $T_{optA}$ , soil soluble phosphorus (P) and potassium (K) were

the only predictors selected for the final model ( $p = 0.004$  and  $p = 0.054$ , respectively; Table S2). However, this relationship can be attributed almost entirely to between-species relationships and not to intraspecific relationships (marginal  $R^2 = 0.09$ , conditional  $R^2 = 0.50$ ). Moreover, in single-species analysis, P had only a marginally significant positive relationship with  $T_{optA}$  in three of the study species, *B. simaruba* ( $p = 0.06$ ), *Q. virginiana* (0.07), and *H. actinophyllum* (0.09). K was not significantly related to  $T_{optA}$  for any single-species analysis.

### 3.3 | Relationships between underlying photosynthetic processes and $T_{optA}$ and $A_{opt}$

$A_{opt}$  correlated positively with  $V_{opt}$ ,  $J_{opt}$  and  $g_{opt}$  for most species (Figure 6d,h,l, Supporting Information: Table S3), but exhibited no relationships with the  $T_{opt}$  of these parameters (Figure 6b,f,j).  $T_{optA}$  consistently correlated with  $T_{optg}$  in all species (Figure 6i, Supporting Information: Table S4), and for some species  $T_{optA}$  correlated with the temperature optima or optimum rates of  $V_{cmax}$ ,  $J_{max}$  or  $g_s$  (Figure 6a,c,e,g,k).

The slope of  $J_{max}$ : $V_{cmax}$  over measurement leaf temperatures was significantly negatively correlated with  $T_{optA}$  for *Q. virginiana* (Figure 7a, Supporting Information: Table S4) and significantly positively correlated with  $A_{opt}$  for *C. uvifera* (Figure 7b, Supporting



**FIGURE 5** Within-species linear least squares relationships between land surface temperature (LST) and (a) the thermal optimum of net photosynthesis ( $T_{optA}$ ) and (b) the rate of photosynthesis at its thermal optimum ( $A_{opt}$ ), and between percent impervious cover within 5 m of each tree and (c)  $T_{optA}$  and (d)  $A_{opt}$  (all nonsignificant,  $p > 0.05$ ). Each point is a single tree.

Information: Figure S3).  $J_{max25} \cdot V_{cmax25}$  was not related to  $T_{optA}$  in any species (Figure 7c, Supporting Information: Table S4) and was significantly negatively associated with  $A_{opt}$  in *B. simaruba* and *T. catappa* (Figure 7d, Supporting Information: Figure S3).  $J_{maxTopt} \cdot V_{cmaxTopt}$  showed a significant negative association with  $T_{optA}$  for *B. simaruba* and *S. mahagoni* (Figure 7e, Supporting Information: Figure S4).  $J_{maxTopt} \cdot V_{cmaxTopt}$  also showed a significant negative relationship with  $A_{opt}$  for *B. simaruba*, *S. mahagoni* and *T. catappa* (Figure 7f, Supporting Information: Figure S3).

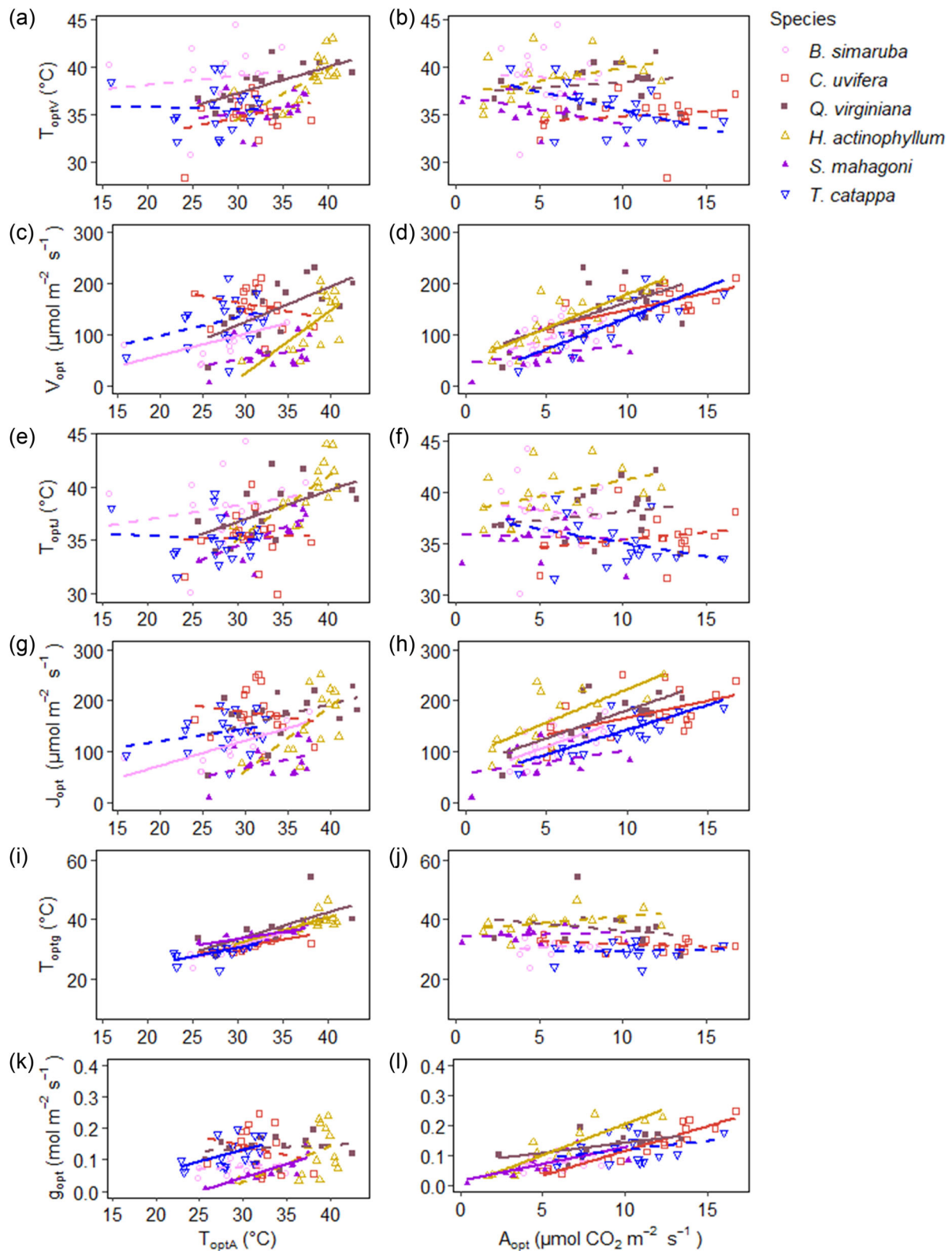
The slope of stomatal limitation ( $l$ ) over leaf temperature decreased significantly with increased  $T_{optA}$  (Figure 7g, Supporting Information: Figure S4) for one species, *Q. virginiana*. In other words, trees of *Q. virginiana* with higher  $T_{optA}$  were less limited by stomatal conductance when leaf temperatures are high, which is consistent with the observation that  $g_s$  and  $A_{net}$  of *Q. virginiana* are comparatively high, even at 40°C.  $l$  showed a significant negative association with  $A_{opt}$  in one species, *H. heptaleurum* (Figure 7h, Supporting Information: Figure S3).

Finally, the individuals of our study species for which the activation state of Rubisco was significantly reduced at the upper end of the temperature response curve (Figure 4—upper tail) did not show a significant association to reduced  $A_{opt}$ .

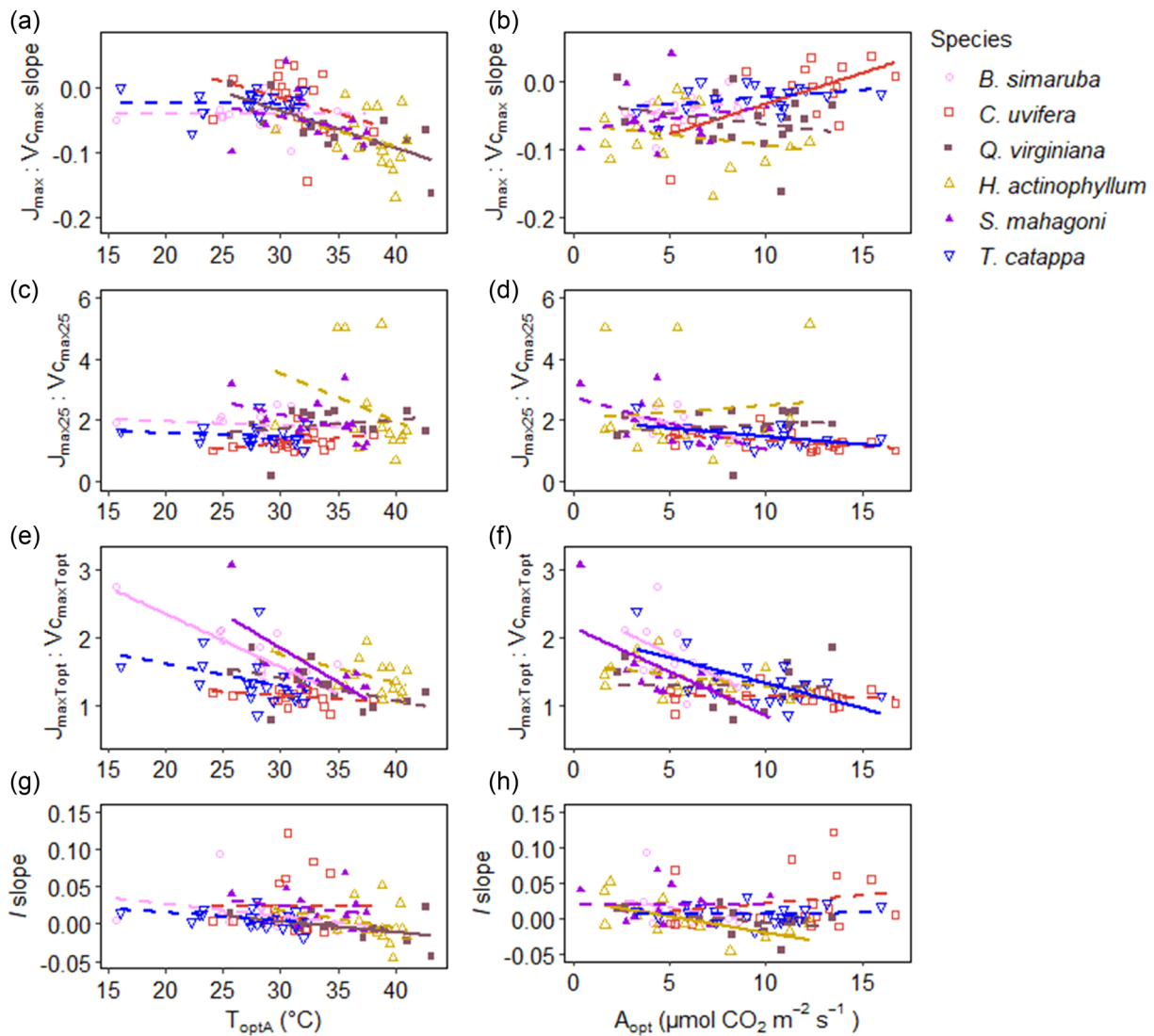
### 3.4 | Limiting factor of $A_{net}$

There were no clear patterns regarding relationships between  $T_{optA}$  and the best predictor of  $A_{net}$  for any of the six study species when using a branch-and-bound algorithm to model  $A_{net}$  (Figure 8a). There were also no major patterns regarding the  $R^2$  of the single-predictor model of  $A_{net}$  and the predictor of the model (i.e., models predicted by any given variable were not necessarily stronger than those predicted by other variables), except for *T. catappa*, which had significantly higher  $R^2$  for models predicted by  $g_s$  compared to those predicted by either  $J_{max}$  ( $p = 0.04$ ) or  $V_{cmax}$  ( $p = 0.002$ ) (Figure 8b). Interestingly, *T. catappa* was also the only species to show a significant LMM relationship between the  $R^2$  of the single-predictor model and  $T_{optA}$  with the model predictor as a random intercept ( $p = 0.007$ , marginal  $R^2 = 0.21$ , conditional  $R^2 = 0.57$ , Supporting Information: Figure S2).

The  $A_{net}$  of most individuals across all six study species was limited by  $g_s$ . The relative frequency of individuals for which  $g_s$  is the single best predictor of  $A_{net}$  across treatment temperatures (0.73, averaged across species) is significantly greater than the frequency of individuals limited by either  $J_{max}$  (mean = 0.09,  $p < 0.0001$ ) or  $V_{cmax}$  (mean = 0.18,  $p < 0.0001$ ) (Figure 8c). The single best predictor of  $A_{net}$



**FIGURE 6** Within-species relationships for the thermal optima ( $T_{opt}$ s) and optimum process rates ( $k_{opt}$ s) of the underlying factors affecting net photosynthesis ( $A_{net}$ ) over  $T_{optA}$  (optimum temperature of net photosynthesis, left (a, c, e, g, i, k) and  $A_{opt}$  (net photosynthetic rate at the optimum temperature, right (b, d, f, h, j, l)). Each point represents an individual tree, solid lines are significant relationships ( $p < 0.05$ ), and dotted lines are nonsignificant relationships.



**FIGURE 7** Within-species relationships for  $T_{\text{optA}}$  (optimum temperature of net photosynthesis, left) and  $A_{\text{opt}}$  (net photosynthetic rate at the optimum temperature, right) explained by (a, b) the slope of  $J_{\text{max}}:V_{\text{cmax}}$  over leaf measurement temperatures ( $T_{\text{leaf}}$ ); (c, d)  $J_{\text{maxTopt}}:V_{\text{cmaxTopt}}$ ; (e, f)  $J_{\text{max25}}:V_{\text{cmax25}}$  and (g, h) the slope of the relationship between stomatal limitation ( $l$ ) (Equations 3 and 4) and  $T_{\text{leaf}}$ . Each point represents an individual tree, solid lines are significant relationships ( $p < 0.05$ ), and dotted lines are nonsignificant relationships.

at high temperatures (above  $T_{\text{optA}}$ ) was the same as when using data from all leaf temperatures in most individuals (77%), and there was no clear pattern for when the best predictor was different between the full model and the high-temperature model.

## 4 | DISCUSSION

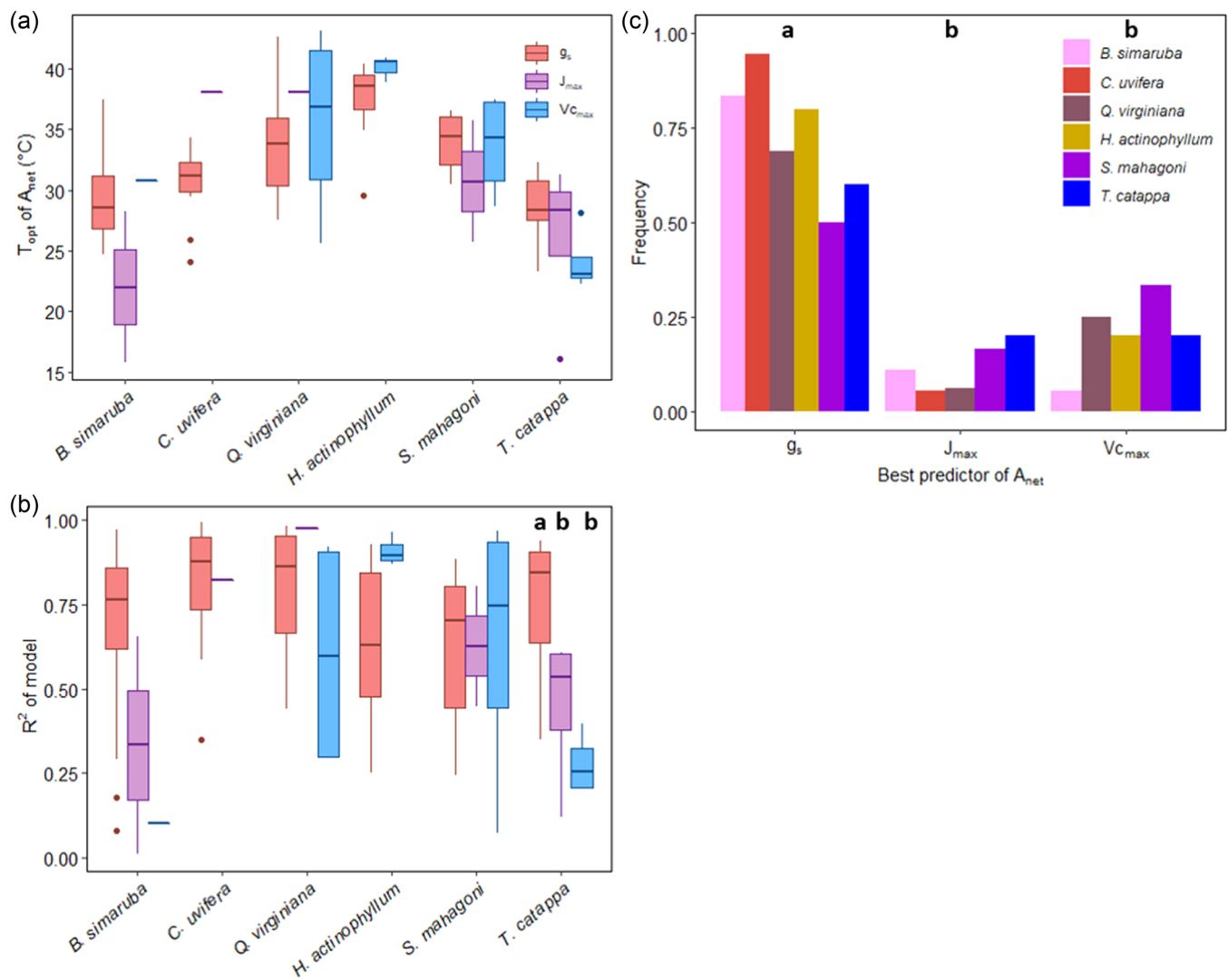
### 4.1 | Photosynthetic thermal optima do not acclimate to different growth temperatures

Contrary to our a priori predictions, the six subtropical tree species that we studied did not acclimate their thermal optima of net photosynthesis ( $T_{\text{optA}}$ ) to temperature differences across the Miami urban heat island. Likewise, although the goodness-of-fit of the

models estimating  $T_{\text{optA}}$  and  $g_s$  were highly variable between individuals, there was little evidence for thermal acclimation for any of the underlying photosynthetic parameters. Notably, against expectations (Crous et al., 2018; Perez et al., 2021), this lack of thermal acclimation did not correspond to significant changes in maximum net photosynthesis rates ( $A_{\text{opt}}$ ) at high growth temperatures. Taken together, these results indicate that growth temperature differences of up to 5°C did not induce thermal acclimation in any of our study species and did not lead to differences in maximum photosynthetic performance.

Our results are similar to those of Dusenge, Wittemann, et al. (2021), who found no evidence of  $T_{\text{optA}}$  acclimation in their study of two tropical montane tree species. However, there is a substantial body of literature that has found significant acclimation of  $T_{\text{optA}}$  to increased growth temperatures (Crous et al., 2022;





**FIGURE 8** (a) The thermal optimum of net photosynthesis ( $T_{optA}$ ) by the photosynthesis component that best predicted  $A_{net}$  for each tree in a single-predictor branch-and-bound algorithm model, separated by study species. (b)  $R^2$  of the single-predictor model of  $A_{net}$  for each tree by the predictor of that model, separated by study species. (c) The relative frequencies of the component of photosynthesis most strongly predicting  $A_{net}$ . Each coloured bar represents individuals of a different species. Letters indicate significant differences between groups (one-way analysis of variance).

Hall et al., 2013; Kumarathunge et al., 2019; Way & Sage, 2008; Yamaguchi et al., 2016), even in tropical tree species (Slot et al., 2021). Contrary to our results, previous studies have also shown decreasing  $A_{opt}$  with increasing  $T_{optA}$  due to performance tradeoffs in the underlying processes (Berry & Bjorkman, 1980; Dusenge, Wittemann, et al., 2021; Slot et al., 2021; Way & Sage, 2008), although in a meta-analysis Way and Yamori (2014) demonstrated that acclimation of  $T_{optA}$  does not correspond to a consistent response in  $A_{opt}$  across 103 species, possibly related to a cross-biome pattern of reduced ability to acclimate towards the tropics (Crous, 2019; Crous et al., 2018).

Although our satellite-derived measures of LST likely provide an accurate characterization of between-site variability in relative growth temperature (Bechtel, 2011; Connors et al., 2013;

Estoque & Murayama, 2017; Kullberg & Feeley, 2022), they may not adequately account for the microclimate conditions of the sample material (Conry et al., 2014; Gluch et al., 2006). Temperatures can vary over spatial scales smaller than 30 m (the resolution of Landsat 8 satellite images), meaning that individual trees or leaves may be acclimated to microclimate conditions that vary considerably from the larger-scale site conditions (Carter et al., 2021). However, an effect of microsite conditions on photosynthetic temperature responses is not always observed (Hernández et al., 2020). In addition, variations of other environmental variables like  $CO_2$  and ozone may obscure a potential temperature effect, although we believe that such effects should be minimized in our study by sampling only within urban Miami and excluding rural or natural areas.

Also, each tree in our study was represented by measurements from a single leaf, which increases the potential for introducing variation that can obscure patterns. To counter this limitation, we only sampled fully sun-exposed leaves, and we sampled trees across a large range of LSTs (maximum of  $\sim 6^{\circ}\text{C}$  between sites) to buffer against the potential effects of microsite differences.

$T_{\text{optA}}$  should acclimate based on leaf temperatures rather than air temperatures (Knoerr & Gay, 1965; Wright et al., 2017). We only sampled fully sun-exposed leaves, and by standardizing the radiation load across sites, patterns in leaf temperatures should follow patterns in LST (our measure of growth temperature) if the trees have sufficient access to water. Nonetheless, acclimation of thermoregulatory traits that impact leaf energy balance, such as leaf area, thickness, spectral properties and stomatal properties, may obscure acclimation of photosynthesis by decoupling leaf temperatures and air temperatures. However, in a previous study, we did not find evidence of strong thermal trait acclimation in these same tree species and urban heat island systems (Kullberg & Feeley, 2022). With recent advances in technology such as drones equipped with thermal cameras, it may be possible to measure in situ canopy leaf temperatures more accurately. Going forward, it will be crucial to utilize these emerging technologies to help characterize the temperature dynamics of urban trees and their physiological responses to climate.

Another possibility for the observed lack of acclimation of  $T_{\text{optA}}$  to growth temperatures is uncertainty in parameter estimation. The parameterization of our  $A_{\text{net}}$  temperature response curves relied on  $\leq 10$  data points to estimate three parameters in a four-parameter non-linear least squares model. The low number of data points compared to estimated parameters meant that the goodness of fit of the  $A_{\text{net}}$  temperature response curves was sometimes low. Inaccurate parameter estimates of  $T_{\text{optA}}$  and  $A_{\text{opt}}$  may obscure relationships with growth temperature.

Finally, the observed lack of  $T_{\text{optA}}$  acclimation across growth temperatures may have been due to adaptational differences between individuals of varying seed sources. The trees we sampled were of various ages and size classes and were located across various municipalities in Miami Dade County, so we anticipate that possible that the sample trees of any given species were most likely from multiple seed sources. However, seed source information was not available for the trees in our study, so we are unable to determine the extent of this potential source of error. Future studies could potentially isolate the effects of acclimation and local adaptation by incorporating information on population genetics.

## 4.2 | Urban heat islands provide a potential window into future climate change effects

Despite the fact that we did not observe the acclimation of photosynthetic parameters to growth temperature, we emphasize the potential utility of the urban heat islands as 'accidental experiments' to test the potential effect of future climate change

on trees and other species (Bussotti et al., 2014; Carreiro & Tripler, 2005; Cregg & Dix, 2001; Farrell et al., 2015). Urban heat islands provide steep thermal gradients with levels of warming on par with predicted climate change for this century and often have planted (i.e., not naturally dispersed) trees that facilitate the separation between adaptive and acclimatory effects—something that is not typically possible in observational studies along large-scale elevation or latitudinal gradients.

## 4.3 | Thermal optima of $V_{\text{cmax}}$ and $J_{\text{max}}$ have moderate control over the thermal optima of $A_{\text{net}}$

Growth temperature and other environmental variables did not significantly impact  $A_{\text{net}}$  within any of our study species. Of all the environmental variables included in this study (LST, impervious cover, soil pH, N, P and K), soil soluble P and K were the only variables that were related to  $T_{\text{optA}}$  according to stepwise LMM analysis. Most of the variability explained by soil P and K, however, was attributable to between-species differences rather than intraspecific variation. Accordingly, for our subsequent analyses, we explored the acclimation of the component processes underlying  $A_{\text{net}}$ , irrespective of abiotic conditions. We found various significant single-species relationships between  $T_{\text{optA}}$  or  $A_{\text{opt}}$  and the  $T_{\text{opt}}$  of some of the individual component processes as well as with their respective optimum process rates ( $k_{\text{opt}}$ ), highlighting the processes that drive  $T_{\text{optA}}$  and  $A_{\text{opt}}$ . Importantly, however, we did not directly measure respiration in the light in this study and were, therefore, unable to determine the relative influence of acclimation of respiration compared to other underlying processes of  $A_{\text{net}}$  on the acclimation of  $T_{\text{optA}}$  and  $A_{\text{opt}}$ .

We found that the  $T_{\text{opt}}$  of both  $V_{\text{cmax}}$  and  $J_{\text{max}}$  were somewhat positively related to  $T_{\text{optA}}$  (observed in two and three species, respectively) and therefore appear to have a moderate role in driving differences in  $T_{\text{optA}}$  across species. This result is consistent with previous studies that have found that these two biochemical thermal responses generally relate to shifts in  $T_{\text{optA}}$  (Crous et al., 2022; Hikosaka et al., 2005; Medlyn et al., 2002; Onoda et al., 2005). In most of our study species (all except for *S. mahagoni*), any observed increase in  $T_{\text{optV}}$  and  $T_{\text{optJ}}$  with  $T_{\text{optA}}$  was not due to increased activation energies,  $H_{\text{aV}}$  and  $H_{\text{aJ}}$  (Dusenge, Wittemann, et al., 2021; Kattge & Knorr, 2007). However, Hikosaka et al. (2005) found  $H_{\text{aV}}$  to be the most important factor allowing the acclimation of  $T_{\text{optA}}$ .

Increased  $T_{\text{optA}}$  was related to decreased entropy terms ( $\Delta S$ ) of the temperature responses of  $V_{\text{cmax}}$  and  $J_{\text{max}}$  in just one species, *H. actinophyllum*, providing only limited support that these intra-specific relationships in our study reflect previous intra- and inter-specific findings that the negative temperature response of  $\Delta S$  is more or less generalized across species (Kattge & Knorr, 2007; Slot et al., 2021). In addition, the activation state of Rubisco in most trees across all the study species did not significantly limit  $A_{\text{net}}$ , and individuals in which Rubisco limitation was observed at high temperatures did not have significantly reduced  $A_{\text{opt}}$ . This result is

similar to some observations on the tropical tree *Tabebuia rosea* (Slot et al., 2021) and various herbaceous species (Sage et al., 2008), but contrary to other results showing significant high-temperature Rubisco deactivation in herbaceous species (Haldimann & Feller, 2005; Yamori et al., 2006) and the boreal tree *Picea mariana* (Sage et al., 2008).

Upregulation of both  $V_{opt}$  and  $J_{opt}$  in most species (four and five species, respectively) was associated with an increase in  $A_{opt}$ , as expected (Crous et al., 2022). Although the thermal optima of  $V_{cmax}$  and  $J_{max}$  play a relatively moderate role in regulating  $T_{optA}$ , the greater the rates of these processes at their respective  $T_{optS}$  ( $V_{opt}$  and  $J_{opt}$ ) still led to significantly increased  $A_{opt}$ .

Finally,  $J_{max} \cdot V_{cmax} \cdot T_{opt}$  was lower when plants had a higher  $T_{optA}$ . This pattern of decreasing  $J_{max} \cdot V_{cmax}$  with increasing  $T_{optA}$  has been found previously (Crous et al., 2022; Dusenge et al., 2020; Hikosaka et al., 2005; Kumarathunge et al., 2019) and is an acclimation response associated with higher growth temperatures, typically associated with raising  $V_{cmax}$  relative to  $J_{max}$  to relieve the carboxylation limitation often observed at high temperatures. Although a clear pattern with growth temperature was not evident in the current study, we did find a significant negative relationship between  $J_{max} \cdot V_{cmax} \cdot T_{opt}$  and  $T_{optA}$  in some species. In other words, when a tree is performing at a relatively low  $T_{optA}$ ,  $J_{max}$  and  $V_{cmax}$  operate relatively lower on their own temperature response curves, where  $J_{max}$  is proportionally greater compared to  $V_{cmax}$  than when the process rates operate closer to their respective thermal optima at higher temperatures. The  $J_{max} \cdot V_{cmax}$  ratio at  $T_{optA}$  can thus vary with  $T_{optA}$  without shifts in the temperature response curves of  $J_{max}$  and  $V_{cmax}$ , and since  $T_{optA}$  was more strongly forced by  $g_s$  than by the two biochemical processes, this appears to explain the pattern observed in our study.

#### 4.4 | $A_{net}$ is most strongly limited by $g_s$ across leaf temperatures

We found multiple lines of evidence suggesting that  $g_s$  is the primary component affecting  $A_{net}$ . While  $V_{cmax}$  and  $J_{max}$  have strong effects on  $A_{net}$  at a given stomatal conductance,  $T_{optA}$  and  $T_{optg}$  are typically lower than  $T_{optV}$  and  $T_{optJ}$ . Thus, between  $T_{optA}$  and  $T_{optV}$  or  $T_{optJ}$ ,  $A_{net}$  decreases with temperature despite the fact that  $V_{cmax}$  and  $J_{max}$  are still increasing—this strongly suggests that at those temperatures,  $A_{net}$  is primarily limited by stomatal conductance to  $CO_2$ . Furthermore,  $T_{optA}$  in all six study species was significantly positively related to  $T_{optg}$  despite considerable variability in the goodness of fit of these two temperature response curves in most species, indicating that the temperature response of  $A_{net}$  is significantly driven by the temperature response of  $g_s$ . However, for trees with higher  $T_{optA}$ , the positive association between  $T_{optA}$  and  $T_{optg}$  was only associated with reduced stomatal limitation ( $l$ ) at high leaf temperatures (i.e., closer to  $T_{optA}$ ) for one species, *Q. virginiana*. In other words, although  $T_{optA}$  is highly dependent on  $T_{optg}$ , trees with a higher  $T_{optA}$  do not necessarily benefit from reduced stomatal limitation when performing at their

$T_{optA}$ , although visually there is a weakly decreasing trend in the slope values of  $l$  over increasing leaf treatment temperatures with increasing  $T_{optA}$ , as was expected. Interestingly, *Q. virginiana* is at the southernmost end of its range in Miami, whereas the other five study species are at their northern range limits, indicating that the plasticity of stomatal limitation may depend on a species' latitude of origin. Like  $J_{max}$  and  $V_{cmax}$ ,  $g_{opt}$  was also positively related to  $A_{opt}$  in four of the study species and showed relatively higher  $R^2$  values than the two biochemical process rates, indicating that the rate of  $g_s$  plays a significant role in limiting  $A_{opt}$ .

Because  $V_{opt}$ ,  $J_{opt}$  and  $g_{opt}$  were all significantly positively correlated with  $A_{opt}$  in most of our study species, it was difficult to determine the overall relative impact of these three processes on  $A_{net}$ . To address this uncertainty, we conducted the single-predictor model selection analysis to determine which of the three processes was the best predictor of  $A_{net}$  across leaf treatment temperatures for each tree. We found that  $g_s$  was the single best predictor of  $A_{net}$  in most trees, which was true across all six species. Out of the six species, *S. mahagoni* had proportionally the fewest individuals in which  $A_{net}$  was best predicted by  $g_s$ , likely because  $g_s$  tended to remain very low across leaf measurement temperatures in this species. The best predictor of  $A_{net}$  was not related to whether the full set of leaf treatment temperatures was included or only those  $>T_{optA}$ , to the  $R^2$  of the model, or to the  $T_{optA}$  of the tree, so we could not determine a reason for the differences in the best predictor between trees of the same species. *T. catappa* was the only exception as the one species to show a significant positive relationship between the  $R^2$  of the single-predictor model and the  $T_{optA}$ . Therefore, for *T. catappa*, individuals with higher  $T_{optA}$  are more strongly limited by a single predictor, and furthermore that predictor is most likely to be  $g_s$ .

There is a growing number of studies on the limitations of  $A_{net}$ , especially in tropical species, that support our finding that stomatal conductance is of great importance (Carter et al., 2021; Grossiord et al., 2020; Rowland et al., 2015; Slot & Winter, 2016, 2017a; M. N. Smith et al., 2020; Tan et al., 2017; Wong et al., 1979). This general finding is in contrast to observations from temperate systems where photosynthesis appears to be more strongly controlled by  $V_{cmax}$  (Sage & Kubien, 2007; Scafaro et al., 2017; Yamaguchi et al., 2016), especially at high temperatures (Hikosaka et al., 2005; Way & Sage, 2008), and is sometimes limited by  $J_{max}$  (Sage & Kubien, 2007). There is, however, recent evidence that even some higher-latitude trees may face stomatal limitation to  $A_{net}$  at elevated temperatures, although the response appears to be species-specific (Dusenge, Ward, et al., 2021).

As the general importance of stomatal conductance in limiting  $A_{net}$  becomes clearer, it is necessary to determine how the confounding effects of temperature and water limitation act to alter rates of stomatal conductance and, in turn, affect  $A_{net}$  (Tan et al., 2017). For example, elevated leaf-to-air VPD, which tends to increase with increased temperatures and decreasing water availability, often causes stomatal closure (Lin et al., 2012). This is especially true in sun versus shade leaves (Hernández et al., 2020) and in tropical versus temperate species (Cunningham, 2004), possibly because of a tropical-to-temperate isohydry-anisohydry

gradient (Meinzer et al., 2017). Moreover, current models suggest that tropical forests are currently operating beyond their thermal and VPD optima (Rowland et al., 2015), so parsing out the specific effects of temperature versus water stress on stomatal conductance is of great importance.

## 5 | CONCLUSION

We did not detect evidence of acclimation of photosynthesis to elevated growth temperatures within adults of six subtropical tree species growing across a range of temperatures in Miami's urban heat island. However, we also did not observe a decline in  $A_{opt}$  at increased growth temperatures, so while there is no evidence for  $T_{optA}$  acclimation, there is no indication that heat island temperatures are limiting the photosynthetic performance of our study species. These results are contrary to the findings of previous studies that have identified limited acclimation of the thermal optimum of net photosynthesis ( $T_{optA}$ ), occasionally at a cost to the rate of net photosynthesis at its thermal optimum ( $A_{opt}$ ). We found evidence that all three underlying photosynthetic components included in our study— $V_{cmax}$ ,  $J_{max}$  and stomatal conductance—have varying levels of control on  $T_{optA}$  as well as  $A_{opt}$ , confirming that they are each at least partially responsible for determining the overall temperature response of  $A_{net}$ . However, the study species differed greatly in terms of correlations among photosynthetic parameters, emphasizing the potential for highly species-specific responses to environment, which complicates the prediction of future tree performance. We also found that  $A_{net}$  was most often limited by stomatal conductance in all six species. This suggests that the controls on  $A_{net}$  in the subtropics are more like those in the tropics than in temperate zones and highlights the need for continued investigation of the environmental controls to stomatal conductance.

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## DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

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## SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

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